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## **Analytical Report**

**PFOA and PFOS Analysis of Deer Muscle and Liver Samples by LC/MS/MS**

**MPI Report No. L0019346**

**Revised Report Date: 12/17/09**

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### ***Testing Laboratory***

MPI Research, Inc.  
3058 Research Drive  
State College, PA 16801

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### ***Requester/Project Manager***

Dena Haverland  
Dalton Utilities  
PO BOX 869  
Dalton, GA 30722  
Phone: 706-529-1010

2010 JAN -6 P 1:44

## 1 Introduction

Results are reported for the analysis of the 3.5 yr male deer liver sample received at MPI Research from Dalton Utilities. The MPI Research study number assigned to the project is L0019346. Table I lists the target analytes quantitated for the samples.

Table I. Target Analytes for Quantitation

Compound Name	Acronym
Perfluorooctanesulfonate	C8 Sulfonate or PFOS

Note: PFOA and PFOS results for all the muscle and liver samples with the exception of the 3.5yr male deer liver sample are reported in the original report signed on 11/19/09.

## 2 Sample Receipt

Four samples were received from Dena Haverland at Dalton Utilities for this study. The samples were collected on October 02, 2009. The samples arrived on October 06, 2009 via Fedex and were logged in under MPI Research login number L0019346. The shipment was received frozen on dry ice. The samples were stored frozen at approximately -20°C from receipt until analysis. Chain-of-custody information is presented in Attachment A.

## 3 Methods - Analytical and Preparatory

### 3.1 Muscle and Liver Sample Preparation

- 3.1.1. Weigh 1 g of muscle or liver sample into a 50 mL disposable centrifuge tube and fortify, if appropriate. Add 20 µL of a 50000 ng/mL WIS for a final concentration of 1 ng/mL.

Note: The internal standard was spiked at a higher level to allow for post extraction dilutions to be performed.

- 3.1.2 Add water to the sample for a final volume of 10 mL. Cap tightly.
- 3.1.3 Homogenize sample using a tissuemizer for ~1 minute.
- 3.1.4 Transfer 1 mL of the sample using a disposable pipette into 15 mL disposable centrifuge tubes. Add 5 mL of ACN and shake for ~20 minutes on a wrist action shaker.
- 3.1.5. Centrifuge tubes at ~3000 rpm for ~ 5 minutes. Carefully decant supernatant into a 50 mL disposable centrifuge tube and add 35 mL of water.
- 3.1.6 Place the unconditioned SPE columns on the vacuum manifold. Condition the SPE columns by passing ~ 10 mL of methanol through the column followed by ~ 5 mL of water. The washes may be pulled through the SPE column using vacuum at a flow rate of ~1 drop/sec or may be allowed to pass through the column unaided. Discard all washes. Do not allow the column to dry.

- 3.1.7 Load the sample onto a conditioned SPE column . Discard the eluate. Any analyte residues will be trapped on the SPE column at this point.
- 3.1.8 Elute with 2 mL of methanol. Collect 2 mL of elute into a graduated 15 mL centrifuge tube.

Note: Post extraction dilutions were prepared in methanol.

### 3.2 Sample Analysis by LC/MS/MS

In High Pressure Liquid Chromatography (HPLC), an aliquot of extract is injected and passed through a liquid-phase chromatographic column. Based on the affinity of the analyte for the stationary phase in the column relative to the liquid mobile phase, the analyte is retained for a characteristic amount of time. Following HPLC separation, mass spectrometry provides a rapid and accurate means for analyzing a wide range of organic compounds. Molecules are ionized, fragmented, and detected. The ions characteristic of the compounds are observed and quantitated against external calibration standards.

An HP1100 system interfaced to an Applied Biosystems API 4000 LC/MS/MS was used to analyze the sample extracts for quantitation. A gradient elution through a Phenomenex Luna 3 $\mu$  C8(2) Mercury, 20 x 4.0 mm column was used for separation.

The following gradient was performed:

Mobile Phase (A): 2mM Ammonium Acetate in Water  
Mobile Phase (B): Methanol

Time	%A	%B
0.0	90	10
0.5	90	10
2.0	10	90
5.0	10	90
5.1	0	100
6.0	0	100
6.1	90	10
10.0	90	10

The following parameters were used for operation of the mass spectrometer:

Parameter	Setting
Ionization Mode	Electrospray
Polarity	Negative
Transitions Monitored	499→80 (PFOS) 503→80 (Internal Std. <sup>13</sup> C PFOS (m+4))
Gas Temperature	450°C

## **4 Analysis by LCMSMS**

### **4.1 Calibration**

For the muscle and liver sample analysis, a 6-point calibration curve was analyzed throughout the analytical sequence for PFOS. The calibration points were prepared at 0.1, 0.2, 0.5, 1.0, 2.0, 5.0 ng/mL (ppb) containing 1.0 ng/mL  $^{13}\text{C}$ -PFOS (m+4).

The ratio of the analyte concentration to the IS concentration versus the ratio of the analyte instrument response (area) to the IS response (area) was plotted for each point. Using linear regression with 1/x weighting, the slope, y-intercept and coefficient of determination ( $r^2$ ) were determined. A calibration curve is acceptable if  $r^2 \geq 0.985$ .

For the results reported here, calibration criteria were met. The calibration curves are included in the raw data in Attachment C.

### **4.2 Laboratory Control Spikes**

Laboratory control spikes in the analytical set were prepared during each extraction set by adding a known concentration of the analyte to deer muscle and liver controls. Laboratory control spikes are used to assess method accuracy. The laboratory control spikes must show recoveries between 70-130% or the data is rejected. For the results reported here, the laboratory control spikes were within the acceptable range. Laboratory control spike recoveries are given in Attachment B.

### **4.3 Matrix Spikes**

A matrix spike was prepared for each sample by adding a known concentration of the target analyte to a sample. Matrix spikes are used to assess method accuracy in the matrix. The matrix spikes should show recoveries between 70-130%. For the results reported here, the matrix spike was within the acceptable range with the exceptions of:

### **4.4 Laboratory Duplicates**

Each sample was prepared in duplicate and analyzed. Duplicate results are given along with the sample results in Attachment B.

## **5 Data Summary**

Please see Attachment B for a detailed listing of the analytical results. For the muscle and liver samples the results are reported in parts per billion (ng/g) on an as-received basis.

## 6 Data/Sample Retention

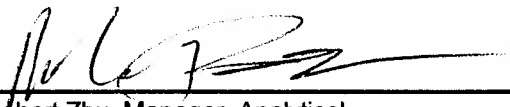
Samples are disposed of 60 days after the report is issued unless otherwise specified by the project manager. All electronic data is archived on retrievable media and hard copy reports are stored in data folders maintained by MPI Research. Hardcopy data is stored for a minimum of five years. The client will be notified 30 days prior to the disposal of hardcopy data.

## 7 Attachments

- 7.1 Attachment A: Chain of Custody
- 7.2 Attachment B: Analytical Results
- 7.3 Attachment C: Raw Analytical Data for Water

## 8 Signatures

  
\_\_\_\_\_  
Mark Neeley, Research Chemist Associate II  
12-17-09  
Date

  
\_\_\_\_\_  
Robert Zhu, Manager, Analytical  
12/17/09  
Date



A







Mattawan (Corporate Headquarters)  
54943 North Main Street  
Mattawan, MI 49071-9399  
(269) 668-3336 Phone  
(269) 668-4151 Fax

State College  
3058 Research Drive  
State College, PA 16801  
(814) 272-1039 Phone  
(814) 231-1580 Fax

## Login

Login Group: L0019346

Login #:	19460	Conform COC Sample:	True
Project:	P0005196	Conform COC:	True
Company Name:	Dalton Utilities	Conform Sample:	True
Submitted By:	Dena Haverland	Conform Request:	True
Login Type:	Immediate Receipt of Samples		
Started:	True		
Date Start:	10/27/2009		
Due Date:	11/06/2009		
Login Initiated:	10/27/2009		
Received By:	Ammerman, Mark		
Spread Sample:			
Label:			
MPI SD/PI:	Zhu, Xiang		
Project Title/Type:	PFOA and PFOS Analysis of Animal Muscle and Liver by LC/MS/MS / ROUTINE		
Login Notes:			

## Packages / Containers

Package	Carton	Date / Condition	Shipper / ID	Temp. Control/Temp.	Direction / Handled By
*K0022042		Received Date: 10/27/09 10:25 Package & Contents Uncompromised	FEDEX 8694 2057 8178	Dry Ice -79.2	RECEIVED Ammerman, Mark

Container #	Gross Weight	pH	Container Type	Preservative	Mfg. Lot	Mfg. ID
C0457624	218.20 g		1/2 gallon ziplock bag	NONE		
C0457625	278.00 g		1/2 gallon ziplock bag	NONE		
C0457626	324.10 g		1 gallon ziploc bag	NONE		
C0457627	994.10 g		1 gallon ziploc bag	NONE		

## Samples

Sample ID	Container	Matrix	System	System Matrix	Sample	Date Sampled	Date Due
L0019346-0001	C0457624	SOLID	Deer	Tissue	Deer #6 0.5 yr female-muscle	10/02/2009	11/06/2009
L0019346-0002	C0457626	SOLID	Deer	Liver	Deer #6 0.5 yr female-liver	10/02/2009	11/06/2009
L0019346-0003	C0457625	SOLID	Deer	Tissue	Deer #7 3.5 yr male-muscle	10/02/2009	11/06/2009
L0019346-0004	C0457627	SOLID	Deer	Liver	Deer #7 3.5 yr male-liver	10/02/2009	11/06/2009



# Login

Login Reviewed By:

[Signature]

Date/Time:

12/16/09 1317



# MPI

RESEARCH

MPI Research Contact: Daniel Wright

## Send Report To:

Company: Dalton Utilities  
 Address: 1200 VD Parrott JR Parkway, PO Box 869  
 City, State, ZIP: Dalton, GA 30722-0869  
 Attention: Dena Haverland  
 Phone #: 706-529-1010  
 Fax #: 706-529-1271  
 Email: dhaverland@dutil.com  
 Study/Job #: \_\_\_\_\_  
 Signature/Date: \_\_\_\_\_  
 Printed Name: \_\_\_\_\_

## Sample Submittal

Please fax this form before sending samples.

Please send samples to shipping and receiving:  
 3048 Research Drive, State College, PA 16801  
 T: (814) 272-1039 • F: (814) 272-1019

## Turnaround time (TAT) requirements:

Results Due Date: 30 days  
 Preliminary Results Format: Verbal ☒ Email ☐ Fax ☐  
 Report Due Date: 30 days

## Storage Conditions

Room temperature  
 Refrigerator  
☒ Freezer  
 Ultra Low freezer  
 Desiccated  
 Lighting required  
 Stability (°C/%RH): \_\_\_\_\_  
 Stability time period: \_\_\_\_\_

## Safety Information

Special handling: \_\_\_\_\_  
 MSDS attached  
 Controlled substance: \_\_\_\_\_  
 HAZARDS \_\_\_\_\_  
 Please fill in the diamond HMIS/NFPA  
 (0-4) if appropriate

Client ID# Description	Lot/ Control #	Amt. Sent/ Weight	# of Bottles	Matrix	Date & Time	Tests Requested
1 <u>Deer #6</u> <u>0.5 yr Female-Serum</u>		<u>10ml</u>	<u>10</u>	<u>deer</u>	<u>10/2/09 1:08am</u>	<u>PFCA/PFOA</u>
2 <u>Deer #6</u> <u>0.5 yr Female-muscle</u>		<u>repacked</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:28am</u>	<u>PFCA/PFOA</u>
3 <u>Deer #6</u> <u>0.5 yr Female-Liver</u>		<u>Whole</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:36am</u>	<u>PFCA/PFOA</u>
4 <u>Deer #7</u> <u>3.5 yr Male-Serum</u>		<u>10ml</u>	<u>10</u>	<u>deer</u>	<u>10/2/09 1:45am</u>	<u>PFCA/PFOA</u>
5 <u>Deer #7</u> <u>3.5 yr Male-muscle</u>		<u>repacked</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:45am</u>	<u>PFCA/PFOA</u>
6 <u>Deer #7</u> <u>3.5 yr Male-Liver</u>		<u>Whole</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:48am</u>	<u>PFCA/PFOA</u>
7						
8						
9						
10						

PO # \_\_\_\_\_

Relinquished by: Daniel Wright Date: 10/5/09 6:55am

Received by: MIA Date: 10/27/09

Notes

"THIS IS AN EXACT COPY OF  
 THE ORIGINAL DOCUMENT"

BY MIA DATE 10/27/09



## TEMPORARY SAMPLE STORAGE FORM

To be completed during ExyLIMS Login

Project #: 15196

Login #: L 15196

Initials / Date: ML 10/27/09

One form to be completed for each package

Date / Time Received: 10/26/09 1025

Received By: Mark Amrhein

Shipper: FedEx

Shipper Package ID: 8694 2057 8178

Temperature (deg C) / Thermometer ID: -79.2 / D0001775

Temperature Control Method: dry ice - active

Temporary Storage Location: walk in freezer 11

Condition of sample(s):

- ☒ Good - Package and contents uncompromised  
☐ Fair - Package damaged / contents uncompromised  
☐ Poor - Package and contents compromised

Notes:

# FedEx® US Airbill

Tracking Number **8694 2057 8J78**

1 From **10/15/04**

Sender's Name **Daniel K. Koonin** Phone **704 998-5537**

Company **USMC 611115 Southfield MI 48111**

Address **2000 Phoenix Road**

City **Ann Arbor** State **MI** ZIP **48106**

## 2 Your Internal Billing Reference

3 To Recipient's Name **Daniel K. Koonin** Phone **704 998-5537**

Company **USMC 611115 Southfield MI 48111**

Recipients Address **2000 Phoenix Road**

City **Ann Arbor** State **MI** ZIP **48106**



8694 2057 8J78

Recipient's Copy

4a Express Package Service

☒ FedEx Priority Overnight ☐ FedEx Standard Overnight

☐ FedEx 2Day ☐ FedEx Express Saver

4b Express Freight Service ☐ FedEx 1Day Freight ☐ FedEx 2Day Freight

5 Packaging ☐ FedEx Pak ☐ FedEx Box ☐ FedEx Tube ☒ Other

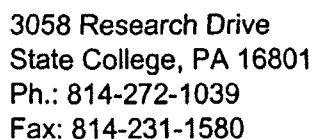
6 Special Handling ☐ No ☐ Yes

7 Payment ☒ Sender ☐ Recipient ☐ Third Party ☐ Credit Card ☐ Cash/Check

8 Residential Delivery Signature Options ☒ No Signature Required ☐ Direct Signature ☐ Indirect Signature

Total Packages **1** Total Weight **1.00** Total Declared Value **\$0.00**

5201



Processing Requested By Employee:	Mark Neeley
MPI Research Assigned Project Number:	P0005196
MPI Research Assigned Login Number:	L0019346

[illegible]

Date Processed: 10/29/2009 Time Started: 14:57 Time Finished: 17:20

Processed By Employee/Employees: Eric Edwards

**Processing Instructions/Comments:**

B







3058 Research Drive  
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## Analytical Report

### Summary of Fluorochemical Residues in Liver Samples

Sample ID	PFOS
	Perfluorooctanesulfonate
	Analyte Found (ng/g, ppb)
Deer # 7 3.5 yr male-liver	2750
Deer # 7 3.5 yr male-liver*	2600

\*Laboratory Duplicate

ND = Not detected = Response is below the LOD of 1.0 ng/g (ppb).

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g (ppb).



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## Recovery Summary of Fluorochemical Residues in Liver Samples

Sample Description	Amount Spiked (ng/g)	Amt Found in Sample (ng/g)	PFOS	
			Amount Recovered (ng/g)	Recovery (%)
LCS A (Data set 112409B) 2000 ng/g	2000	16.5	2340	116
LCS B (Data set 112409B) 2000 ng/g	2000	16.5	2170	108
Deer # 7 3.5 yr male-liver (L19346-4 Spk C, 2000 ng/g Lab Spike)	2000	2750	4770	101

ND = Not detected = Response is below the LOD of 1.0 ng/g.

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g.